

NEW RT-PCR FOR CLASSICAL SWINE FEVER VIRUS DETECTION IN SWINE AND WILD BOAR, ALLOWING FOR PARALLEL TESTING WITH ID GENE ASF qPCRs

Léa DESPOIS¹, Loic SAVARIT¹, Emilie BIANCHINI¹, Adrien LIMOZIN¹, Anna GREATREX¹, Loic COMTET¹, Philippe POURQUIER¹ ¹ IDvet - Innovative Diagnostics, Grabels, France

INTRODUCTION

CONCLUSION

The ID Gene[™] Classical swine fever virus Duplex kit offers:

Classical swine fever (CSF) is a contagious viral disease of pigs, including wild boar. The causativevirus is a member of the genus Pestivirus of the family Flaviviridae, and is closely related to the viruses of bovine viral diarrhoea and border disease. There is only one serotype of CSF virus (CSFV). Reliable and accurate diagnostics are the key for rapid implementation of control measures to detect and prevent the spread of CSFV.

Innovative Diagnostics has developed a rapid and specific RT-qPCR for detection of CSFV RNA, the ID GENE[™] Classical Swine Fever Virus duplex (product code: IDCSF).

KIT PRINCIPLE

→ Excellent performance: high inclusivity on all 21 CSFV strains; LDPCR = 4 copies / PCR → Optimal test reliability thanks to swine cells specific endogenous internal control included in the kit

→ Rapid and specific detection of RNA from CSFV (target in the non 5' UTR region)

3

→ Can be performed in one cycler run with ID Gene[™] African swine ever qPCR kits, allowing, from the same extracts, for a parallel testing CSFV and ASFV, which share similar clinical patterns

The ID GeneTM Classical Swine Fever Duplex kit is a ready-to-use RT-qPCR kit assay detecting simultaneously CSFV and an endogenous internal positive control.

				ENDOGEN		
CHARACTERISTICS		SAMPLE CSFV	CSFV	CSFV OUS	INTERPRETATION	ID Gene™ Classical Swine Fever Virus duples
Targets	CSFV and endogenous control	RESULT	SIGNAL	SIGNAL		
Matrices	Blood, serum, swabs, Organ and tissues	Positive	+	+/-	Valid; Animal detected as positive for CSFV No signal for endogenous because of competition	
Indiv	ndividual samples or pools up to 10 or 20	Negative		+	Valid; Animal detected as negative for CSFV	
Duration	60 min (rapid protocol)				Not Valid; Analytical process successful, no signal for	V.
Species	Domestic pig, wild boar, warthogs	Degraded			endogenous control due to poor sample quality	
Format	Ready-to-use liquid format	sample or partial			A problem occurred during sample distribution or extraction processes and/or RT-gPCR reaction was	
		PCR inhibition			inhibited. Dilute the extracted DNA 10 times in nuclease-free water	
					or re-extract the sample or consider it uninterpretable.	J

RESULTS

INCLUSIVITY

Analysis of 32 reference CSFV strains provided by the Friedrich-Loeffler-Institut (FLI, Germany) and the French National Reference Laboratory (ANSES Ploufragan, France).

CSFV STRAIN	SUB- GENOTYPE	ORIGIN	IDCSFV RESULTS	
Baker A	1.2		Detected	
Guatemala HC	1.3		Detected	
VRI 4167	1.3		Detected	
39 Margarita	1.4		Detected	
PR VP32/10	1.4		Detected	
907/1	2.1		Detected	
A-2	2.1		Detected	\frown
Parma 98	2.2		Detected	(\checkmark)
Bergen	2,.2	FLI, Germany	Detected	Č,
100/06	2.3		Detected	0
Kanagawa	3.4		Detected	
C strain	1.1		Detected	The ID Course
Stamm	1.1		Detected	The ID Gene
Koslov1128	1.2		Detected	
Schweizll	2.1		Detected	CSF QPCK KIT
Pader	2.1		Detected	
Israel	2.1		Detected	successfully
Bergen	2.2		Detected	
D4886/82/Ro	2.2		Detected	detected all
Uelzen	2.3		Detected	to dealer an end
Spante	2.3		Detected	isolates tested
Congenital Tremor	3.1		Detected	
Kanagawa	3.4		Detected	
Alfort	1.1		Detected	
Brescia	1.2		Detected	
Diepholz	2.3		Detected	
Roesrath	2.3		Detected	
Visbeck	3.1		Detected	
Moselle1	2.3	ANSES, France	Detected	
Bas rhin	2.3		Detected	
Peru I8	1.1		Detected	
Paderborn	2.1		Detected	
Litauen	2.1		Detected	
Evstrup	1.1		Detected	

EXCLUSIVITY

The following 69 selected isolates were tested:

SAMPLE	STRAINS	IDCSFV		
NUMBER	STRAINS	RESULTS		
1-4	Influenza (H1N1, H5N2, H7N1,			
	H9N2)	Not detected		
5-11	African Swine fever	Not detected	The ID Gene CSF aPCR	
12-13	PRRSV (EU and NA types)	Not detected	kit did not show any	
14-24	BVDV (1a, 1c, 1d, 1e, 1g, 1h,	Not detected	cross-reactions with	
	1l, 2a, 2b, 3, 4)	Not detected		
25-28	BHV (1,2,3,4)	Not detected	pathogens tested	
	40 other viruses, bacteria and			
29-69	parasites from different	Not detected		
	species			

OVERALL PERFORMANCE

PCR CHARACTERISTICS			
Efficiency / R ²	94,09 % / 0,99		
Limit of detection	4 copies/PCR with rapid amplification program		
Robustness	Unaffected by all parameters tested (temperature and volume of sample)		
METHOD CHARACTERISTICS (WITH THE ID GENE™ MAG FAST			
EXTRACTION KIT)			
Experimental limit of detection	3,2 x10 ³ copies/ml for swine blood, serum		
(with rapid amplification program	nj and spieen samples		

DIAGNOSTIC SENSITIVITY AND SPECIFICITY

Diagnostic sensitivity and specificity was evaluated on 66 field samples provided by ANSES Ploufragan (France), Landeslabor Schleswig-Holstein Neumünster (Germany) and the FLI (Germany) previously characterized by other methods.

SAMPLE TYPE	NUMBER OF SAMPLES TESTED	DIAGNOSTIC SENSITIVITY (Se) CSFV T	DIAGNOSTIC SPECIFICITY (Sp) ARGET
Organs or tissues	11	4/4	7/7
Whole bloods and serums	22	10/10	12/12
Swabs	23	N/A*	23/23
Cellular culture supernatants	10	7/7	3/3
TOTAL	66	21/21 100%	45/45 100%
* N/A : Non analyzed			

The test correctly identified all positive and negative samples tested

(1) OIE Terrestrial Manual, Classical Swine Fever (infection with classical swine fever virus), chapter 3.8.3, 2019.

COMPARISON WITH ANOTHER COMMERCIALLY AVAILABLE RT-qPCR KIT (KIT A)

The performances of the ID Gene CSF test were compared to another commercially available RT-qPCR kit (Kit A) on 21 RNAs provided by the FLI (Hannover, Germany) and 9 RNAs from EILA panel (Ring test trial 2018) composed of field samples spiked with different CSFV strains provided by ANSES (Ploufragan – France). All samples were tested with the Kit A and the ID Gene assay.



Innovative Diagnostics, 310 rue Louis Pasteur, 34790 Grabels - France • www.innovative-diagnostics.com • info@innovative-diagnostics.com